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Relationship between (non)linear phase II pulmonary oxygen uptake kinetics with skeletal muscle oxygenation and age in 11 to 15 y olds

Breese, Brynmor

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1 **RELATIONSHIP BETWEEN (NON)LINEAR PHASE II PULMONARY OXYGEN UPTAKE**
2 **KINETICS WITH SKELETAL MUSCLE OXYGENATION AND AGE IN 11 TO 15 Y OLDS**

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4 ¹Brynmor C. Breese, ²Zoe L. Saynor, ³Alan R. Barker, ³Neil Armstrong and ³Craig A.
5 **Williams**

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7 **Running Head:** Oxygen uptake kinetics in youth

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9 *¹School of Biomedical Sciences, Faculty of Medicine and Dentistry, University of Plymouth,*
10 *UK; ²Department of Sport and Exercise Science, Faculty of Science, University of*
11 *Portsmouth, UK; ³Children's Health and Exercise Research Centre, Sport and Health*
12 *Sciences, College of Life and Environmental Sciences, University of Exeter, UK.*

13
14 Correspondence:

15 Prof. Craig A Williams, Ph.D.

16 Children's Health and Exercise Research Centre, Sport and Health Sciences, University of
17 Exeter, St Luke's Campus, Heavitree Road, Exeter, EX1 2LU, UK.

18 Email: C.A.Williams@exeter.ac.uk

19 Tel: (+44) 01392 264890

20 Fax: (+44) 01392 724726

New Findings

- **What is the central question of this study?**

To investigate if the phase II parameters of pulmonary oxygen uptake ($\dot{V}O_2$) kinetics display linear, first-order behavior in association with alterations in skeletal muscle oxygenation during step cycling of different intensities or when exercise is initiated from an elevated work rate in youth.

- **What is the main finding and its importance?**

We demonstrate how both linear and non-linear features of phase II $\dot{V}O_2$ kinetics may be determined by alterations in the dynamic balance between microvascular O_2 delivery/utilization in 11 to 15 y olds. We further implicate how the recruitment of higher-order (i.e. type II) muscle fibers during “work-to-work” cycling might be responsible for modulating $\dot{V}O_2$ kinetics with chronological age.

ABSTRACT

This study investigated in nineteen male youth (mean age: 13.6 ± 1.1 y, range: 11.7 – 15.7 y) the relationship between pulmonary oxygen uptake ($\dot{V}O_2$) and muscle deoxygenation kinetics during moderate- and very heavy-intensity ‘step’ cycling initiated from unloaded pedaling (i.e. U→M and U→VH) and moderate-to-very heavy-intensity step cycling (i.e. M→VH). Pulmonary $\dot{V}O_2$ was measured breath-by-breath and tissue oxygenation index (TOI) of the vastus lateralis using near-infrared spectroscopy. There were no significant differences in the phase II time constant ($\tau\dot{V}O_{2p}$) between U→M and U→VH (23 ± 6 s vs. 25 ± 7 s; $P = 0.36$); however, the $\tau\dot{V}O_{2p}$ was slower during M→VH (42 ± 16 s) compared to other conditions ($P < 0.001$). Quadriceps TOI decreased with a faster ($P < 0.01$) mean response time (MRT ; i.e. time delay + τ) during U→VH (14 ± 2 s) compared to U→M (22 ± 4 s) and M→VH (20 ± 6 s). The difference (Δ) between the $\tau\dot{V}O_{2p}$ and MRT -TOI was greater during U→VH compared to U→M (12 ± 7 vs. 2 ± 7 s, $P < 0.001$) and during M→VH (23 ± 15 s) compared to other conditions ($P < 0.02$), suggesting an increased proportional speeding of fractional O_2 extraction. The slowing of the $\tau\dot{V}O_{2p}$ during M→VH relative to U→M and U→VH correlated positively with chronological age ($r = 0.68$ and 0.57 , respectively, $P < 0.01$). In youth, “work-to-work” transitions slowed microvascular O_2 delivery-to- O_2 utilization with alterations in phase II $\dot{V}O_2$ dynamics accentuated between the ages of 11 to 15 y.

Keywords: oxygen uptake time constant, microvascular blood flow, oxygen utilization, near-infrared spectroscopy, muscle fiber recruitment, youth

INTRODUCTION

Following the onset of step exercise, the time constant of phase II pulmonary oxygen uptake (i.e. $\tau\dot{V}O_{2p}$) coheres with that observed for muscle $\dot{V}O_2$ kinetics (Grassi *et al.*, 1996; Krustrup *et al.*, 2009; Benson *et al.*, 2013), or, as its surrogate, phosphocreatine (PCr) breakdown in adults (Rossiter *et al.*, 1999) and children (Barker *et al.*, 2008). However, whilst a progressive slowing of the $\tau\dot{V}O_{2p}$ in older adults (Babcock *et al.*, 1994; DeLorey *et al.*, 2005) has been reported to originate during childhood (see McNarry, 2019 for a recent review), the physiological factors limiting $\dot{V}O_2$ kinetics remain less well understood in youth.

A first-order rate reaction controlling $\dot{V}O_2$ kinetics mandates that the response parameters obey the law of superimposition (Fujihara *et al.*, 1973a; Fujihara *et al.*, 1973b). That is, the $\tau\dot{V}O_{2p}$ and primary gain (G_p ; expressed as the $\dot{V}O_2$ per unit increment in work rate) remain constant following the onset of exercise of different intensities. In adults, whilst a slower $\tau\dot{V}O_{2p}$ has been reported during step exercise above the lactate threshold (>LT) compared to <LT (Paterson & Whipp, 1991; Koppo *et al.*, 2004; McNarry *et al.*, 2012), other studies have reported no significant differences (Ozyener *et al.*, 2001; Wilkerson *et al.*, 2004). A slower $\tau\dot{V}O_{2p}$ has been interpreted by some authors to reflect slower O_2 transport during supra-LT transitions (Hughson *et al.*, 2001; McNarry *et al.*, 2012). Conversely, in children, an invariant $\tau\dot{V}O_{2p}$ during step exercise at progressively higher work rates (Hebestreit *et al.*, 1998; Williams *et al.*, 2001) or following “priming” exercise (Barker *et al.*, 2010; Barker *et al.*, 2014) suggests their phase II $\dot{V}O_2$ kinetics are principally limited by intracellular metabolic factors. However, in youth, the possibility that O_2 delivery might constrain the $\tau\dot{V}O_{2p}$ in an exercise intensity dependent manner

has previously relied on measures such as heart rate dynamics (Hebestreit *et al.*, 1998; Breese *et al.*, 2012), which, are removed from peripheral sites of O₂ exchange between the capillary and muscle.

The τ of muscle deoxyhemoglobin/myoglobin (deoxy[Hb+Mb]) measured by near-infrared spectroscopy (NIRS) has been reported to cohere with that of fractional O₂ extraction (Koga *et al.*, 2012), hence, has been used to reflect the dynamic matching between O₂ delivery- (\dot{Q}_{O_2}) to- O₂ utilization (\dot{V}_{O_2}) during exercise (DeLorey *et al.*, 2003; Grassi *et al.*, 2003). Accordingly, for the same \dot{V}_{O_2} kinetics, an enhanced $\dot{Q}_{O_2}/\dot{V}_{O_2}$ response would be expected to slow deoxy[Hb+Mb] dynamics, whereas, slower \dot{V}_{O_2} kinetics alongside a faster deoxy[Hb+Mb] mean response time (MRT; i.e. time delay + τ) has been interpreted to reflect limited microvascular O₂ delivery during the on-transition of exercise (Murias *et al.*, 2011; Spencer *et al.*, 2012; Murias *et al.*, 2014). Therefore, if, based on adults studies, the kinetics of bulk O₂ delivery were slower during heavy- (>LT) compared to moderate-intensity (<LT) step transitions (Koga *et al.*, 2005; McNarry *et al.*, 2012), an enhanced muscle oxidative capacity in children (Ratel *et al.*, 2008; Tonson *et al.*, 2010) may serve to maintain linearity of their $\tau\dot{V}_{O_{2p}}$ by speeding fractional O₂ extraction during supra-LT transitions in youth.

Dynamic non-linearity with respect to an increased $\tau\dot{V}_{O_{2p}}$ and G_p has also been reported when initiating cycling transitions from an elevated work rate (Hughson & Morrissey, 1982; Brittain *et al.*, 2001; Wilkerson & Jones, 2006, 2007), with these effects suggested to reflect the recruitment of higher-order (i.e. type II) muscle fibers (Brittain *et al.*, 2001; Wilkerson & Jones, 2006, 2007); however, other factors have been implicated (DiMenna *et al.*, 2010a; Bowen *et al.*, 2011; Wust *et al.*, 2014). *In vitro*, type

II muscle fibers display slower $\dot{V}O_2$ kinetics and an increased ATP cost of force production compared to type I muscle fibers (Crow & Kushmerick, 1982). In this regard, a previous study has reported conversion of type I-to-II muscle fibers within the vastus lateralis between the ages of 5 to 20 y (Lexell *et al.*, 1992) with longitudinal alterations in children's $\dot{V}O_2$ kinetics (Fawcner & Armstrong, 2004; Breese *et al.*, 2010) showing commonality with the $\dot{V}O_2$ profiles previously reported in adults with an increased distribution of type II muscle fibers (Barstow *et al.*, 1996; Pringle *et al.*, 2003). Therefore, whilst a slower $\tau\dot{V}O_{2p}$ has been reported during "work-to-work" cycling in 11 to 13 y olds (Breese *et al.*, 2012), whether effects on phase II $\dot{V}O_2$ kinetics might be amplified with increased chronological age is unclear. Additionally, whilst, previous reports of a slower τPCr during work-to-work exercise supports an intrinsic slowness of O_2 utilization in adults (Jones *et al.*, 2008; DiMenna *et al.*, 2010b), this proposal has not been investigated in youth in whom measurement of deoxy[Hb+Mb] responses would provide mechanistic insight by serving as a proxy for muscle fractional O_2 extraction.

Therefore, the primary purpose of this study was to investigate whether phase II $\dot{V}O_2$ kinetics display first-order, linear behavior in association with alterations in deoxy[Hb+Mb] kinetics in 11 to 15 y old boys. We hypothesized that a constant $\tau\dot{V}O_{2p}$ during very heavy- compared to moderate-intensity cycling transitions elicited from unloaded pedaling (i.e. U→VH vs. U→M) would coincide with a faster deoxy[Hb+Mb] MRT, whereas, moderate-to-very heavy-intensity cycling transitions (i.e. M→VH) would slow the $\tau\dot{V}O_{2p}$ alongside a slower deoxy[Hb+Mb] MRT compared to other conditions. Finally, we hypothesized that an increased $\tau\dot{V}O_{2p}$ and G_p following the onset of M→VH would correlate positively with chronological age.

METHODS

Ethical Approval

Prior to participation, rights to confidentiality, withdrawal and benefits/risks of the study were explained with fully informed written assent and consent obtained from each participant and their parent(s) / guardian(s), respectively. All experimental procedures were approved by the Sport and Health Sciences research ethics committee at the University of Exeter (7-5-08#4) and conform to the standards set forth by the *Declaration of Helsinki*, except for registration in a database.

Participants

Nineteen boys (mean \pm SD age: 13.6 ± 1.1 y, range: 11.7 – 15.7 y; stature: 160 ± 13 cm; and body mass: 47.9 ± 11.3 kg) volunteered to participate in this study. The data for 8/19 children were included from a previous investigation (Breese *et al.*, 2012) using the same experimental procedures described below. The participants y from peak height velocity (PHV) was used as a descriptor of somatic maturity level using age and sitting height in a validated algorithm in male youth (Moore *et al.*, 2015). This analysis revealed that ten participants were less than or equal to – 1 y from (i.e., pre-) PHV, with five at PHV, and four greater than 1 y from (i.e. post-) PHV, respectively.

Experimental protocol

Participants attended the laboratory on five to nine occasions over a two to four week period with each visit separated by ≥ 48 h. All cycling tests were performed on an electronically-braked cycle ergometer (Lode Excalibur Sport, Groningen, the

Netherlands) with the seat, handlebar height, and crank length adjusted for each participant and subsequently maintained for all visits. All participants were asked to arrive at the laboratory at least 2 h postprandial and having refrained from caffeine for > 2 h.

On their first visit, each participant performed a ramp incremental cycle test until task failure for determination of their peak $\dot{V}O_2$ and the gas exchange threshold (GET). Following 3-min baseline cycling at 15 W, the work rate increased continuously by 15 W/min in 11 to 13 y olds and 25 W/min in all other participants based on the ramp rates previously estimated to attain a test duration of $\sim 8 - 12$ min across similar age categories (Fawkner & Armstrong, 2004; Breese *et al.*, 2010). Participants were instructed to maintain a pedal rate of 70-80 rpm throughout the test with exhaustion defined as a ≥ 10 rpm drop in cadence for five consecutive seconds despite strong verbal encouragement. The peak $\dot{V}O_2$ was taken as the highest 10-s stationary average value during the ramp test which has been shown previously to reflect a maximum $\dot{V}O_2$ in $\sim 93\%$ of youth performing ramp cycling (Barker *et al.*, 2011; Sansum *et al.*, 2019). The GET was determined using the V-slope method (Beaver *et al.*, 1986) as the first disproportionate increase in CO_2 production ($\dot{V}CO_2$) relative to the increase in $\dot{V}O_2$, and subsequently verified from visual inspection of the increase in the ventilatory equivalent for $\dot{V}O_2$ ($\dot{V}_E/\dot{V}O_2$) with no increase in $\dot{V}_E/\dot{V}CO_2$.

The cycling work rates corresponding to 90% GET and 60% of the difference (Δ) between the GET and peak $\dot{V}O_2$ were estimated using the “linear” portion of the ramp test by removing the initial 2 and final 3 min of test data and following adjustment of the $\dot{V}O_2$ “lag time” during ramp exercise (Whipp *et al.*, 1981). This yielded mean cycling

work rates of 72 ± 22 W equivalent to 90% GET (i.e. moderate-intensity cycling) and ± 38 W equivalent to $\Delta 60\%$ (i.e. very heavy-intensity cycling). Each participant then returned to the laboratory to perform 1 of 2 step exercise protocols consisting of: 1) 3-min cycling at 15 W followed by 6-min of very heavy-intensity cycling (U→VH); or, 2) 3-min cycling at 15 W, followed by 4-min of moderate-intensity cycling (U→M), and then 6-min of very heavy-intensity cycling (M→VH). Each participant completed a minimum of two transitions within each step condition presented in random order.

Experimental measures

Pulmonary gas exchange and ventilation were measured and displayed breath-by-breath during each cycling trial (Metalyser 3B Cortex, Biophysik, Leipzig, Germany). Expiratory and inspiratory flows and volumes were measured via a pediatric facemask with low dead space (~ 45 ml) connected to a low-resistance (≤ 0.1 kPa/l/s at 20 l/s) digital turbine volume transducer which was manually calibrated using a 3-liter syringe (Hans Rudolph, Kansas City, MO) before each exercise test. Respired gases were continuously sampled from the facemask and analyzed for relative concentrations using an electrochemical oxygen sensor with a response time of < 100 ms. The delay in the capillary gas transit and analyzer rise time were accounted for relative to the volume signal, thereby time aligning the concentration and volume signals. Heart rate (HR) was recorded every breath during all cycling tests using short-range telemetry (Polar S610, Polar Electro Oy, Kempele, Finland).

A portable continuous wave (CW-) NIRS device (Portamon, Artnis Medical Systems, the Netherlands) was used to assess skeletal muscle oxygenation of the vastus lateralis

by emitting photons at two separate wavelengths (760 and 850 nm). The sampling frequency was set at 10 Hz. The spacing between the photon emitter and detector was 3.5 cm, corresponding to a depth resolution of 1.5 – 2 cm. The NIRS probe was affixed midway between the greater trochanter and lateral epicondyle of the femur using physiotherapists tape (Kinesio Tex Gold), and secured by an elastic bandage to ensure the device remained stationary and to eliminate contamination from ambient light, thereby, improving the signal-to-noise ratio.

The instrument employed a modified Beer Lambert law to estimate in micromolar (μM) concentration changes in oxygenated and deoxygenated hemoglobin and myoglobin (i.e. $\Delta\text{oxy}[\text{Hb}+\text{Mb}]$ and $\Delta\text{deoxy}[\text{Hb}+\text{Mb}]$) with respect to an initial resting value arbitrarily set equal to zero. A differential path-length factor (DPF) of 4 cm was employed to account for tissue scattering. Since assuming a constant DPF using CW-NIRS cannot resolve absolute $[\text{Hb}+\text{Mb}]$ concentrations (Barstow, 2019), the $\Delta\text{deoxy}[\text{Hb}+\text{Mb}]$ amplitude was normalized relative to the end-exercise value prior to kinetic analysis in each condition. The tissue oxygenation index (TOI; $\text{oxy}[\text{Hb}+\text{Mb}]/\text{oxy}[\text{Hb}+\text{Mb}] + \text{deoxy}[\text{Hb}+\text{Mb}]$, expressed as a percentage) was also calculated by spatially resolved spectroscopy as the TOI is thought to be less sensitive to changes in microvascular volume than $\text{deoxy}[\text{Hb}+\text{Mb}]$ data (Quaresima & Ferrari, 2009).

Data analysis and kinetic modeling

The breath-by-breath $\dot{V}\text{O}_2$ data from each step transition were initially edited to exclude errant breaths by removing values lying more than four standard deviations from the local mean determined using a 5-breath rolling average. The filtered $\dot{V}\text{O}_2$ and

deoxy[Hb+Mb] responses were subsequently linearly interpolated with identical repetitions of each step condition time aligned to the start of exercise and ensemble averaged to improve the signal-to-noise ratio.

The first 15 s of $\dot{V}O_2$ data after the onset of exercise was deleted to remove the phase I (cardio-dynamic) response, and a mono-exponential model with time delay was then fitted to the averaged $\dot{V}O_2$ data of the following form:

$$\Delta Y_{(t)} = \Delta Y_p \cdot (1 - e^{-(t-TD)/\tau_p}) \quad (1)$$

where $\Delta Y_{(t)}$ indicates the value at a given time (t) minus the baseline value (60-s average) before exercise onset, ΔY_p indicates the amplitude change of the primary component from baseline to its asymptote, TD and τ_p represent the time delay and time constant of the phase II exponential function, respectively. For U→M, the model in Equation (1) was fitted to end-exercise (i.e. 4-min), whereas, for U→VH and M→VH, the model fitting window was constrained to exclude the A_{Sc} and hence isolate the phase II component. The onset of the A_{Sc} was determined using software (LabView, v 6.1, National Instruments, Newbury, UK) which initially fitted a mono-exponential function up to the first 60-s of $\dot{V}O_2$ data and then increased iteratively by 5-s until end-exercise. The estimated τ for each fitting window was then plotted against time with the phase II portion of the response determined as the point at which the influence of the A_{Sc} lengthened the estimated τ following an initial plateau (Rossiter *et al.*, 2001). The parameter estimates from Equation (1) and their 95% confidence intervals (CI₉₅) were then resolved by least-squares non-linear regression (GraphPad Prism, GraphPad

Software, San Diego, CA). The A_{Sc} was subsequently determined by calculating the difference between the end-exercise $\dot{V}O_2$ and the sum of the primary amplitude and baseline $\dot{V}O_2$. For all conditions, the ‘gain’ of the phase II response (G_p) was calculated by dividing the asymptotic phase II amplitude minus the baseline $\dot{V}O_2$ by the increment in work rate ($\Delta\dot{V}O_2/\Delta W$). Likewise, the total $\dot{V}O_2$ gain (G_{tot}) at end-exercise was calculated in a similar manner.

The NIRS-derived deoxy[Hb+Mb] and TOI response were also modelled to provide information on the kinetic adjustment of fractional O_2 extraction. The TD for an exponential-like rise in muscle deoxygenation was defined as the first datum lying > 1 SD above the mean value during baseline cycling as previously described (DeLorey *et al.*, 2003). Subsequently, following removal of data points preceding the TD, the model in Equation (1) was fitted to the initial 90 – 120 s of data to resolve the $\tau\Delta\text{deoxy[Hb+Mb]}$ and τTOI , or, in cases where visual inspection revealed an early ‘overshoot’ in muscle deoxygenation relative to end-exercise, to the peak value attained during the transient phase. Finally, the TD and τ were summed to reflect the overall mean response time (MRT) of $\Delta\text{deoxy[Hb+Mb]}$ and TOI within each step condition.

The ratio of $\Delta\text{deoxy[Hb+Mb]}$ to $\dot{V}O_2$ was also calculated using the methods originally described in adults (Murias *et al.*, 2010) and subsequently in children (Barker *et al.*, 2014), to infer the dynamic matching of $\dot{Q}O_2$ -to- $\dot{V}O_2$ during step cycling. Briefly, the $\Delta\text{deoxy[Hb+Mb]}$ and $\dot{V}O_2$ profiles were normalized such that 0% and 100% represented the values corresponding to baseline and at end-exercise, respectively. Subsequently, the $\Delta\text{deoxy[Hb+Mb]}$ and $\Delta\dot{V}O_2$ data were averaged into 5 s bins and time aligned by left shifting the $\dot{V}O_2$ data by 15 s to account for the duration of phase I estimated previously

in children (Springer *et al.*, 1991; Hebestreit *et al.*, 1998). The magnitude of the $\Delta\text{deoxy[Hb+Mb]}/\Delta\dot{V}\text{O}_2$ “overshoot” was calculated by integrating the area under curve from the first datum lying above 1.0 or ‘unity’ to 180-s of exercise in all participants in each condition.

Statistical Analysis

Gaussian distribution was assessed by the Shapiro-Wilk test and subsequently verified by calculating standardized scores for skewness and kurtosis for each variable. A standardized value < 2 was deemed acceptably normally distributed. All pulmonary $\dot{V}\text{O}_2$ and NIRS-derived variables were analyzed using one-way repeated measures ANOVA with Bonferroni adjusted post hoc tests used to locate statistically significant differences between step conditions. In addition, effect size (ES; using Cohen’s *d*) was also calculated to judge the magnitude of the observed effect, using the following thresholds: Trivial (< 0.2), Small (0.2), Medium (0.5), and Large (0.8). Pearson product moment correlations (*r*) were used to assess the bivariate relationship between alterations in phase II $\dot{V}\text{O}_2$ kinetics with muscle oxygenation and chronological age. All statistical analyses were conducted using PASW Statistics 18 (SPSS, Chicago, IL). Data are presented as means \pm SD. Statistical significance was accepted if $P < 0.05$.

RESULTS

The group mean \pm SD values for peak $\dot{V}\text{O}_2$ and end HR during the initial ramp incremental cycle test were 2.37 ± 0.60 l/min and 192 ± 9 bpm, respectively. The group

mean \pm SD values for end HR during U \rightarrow M, U \rightarrow VH and M \rightarrow VH step cycling were 129 ± 16 , 178 ± 11 , and 179 ± 12 bpm, respectively.

Pulmonary $\dot{V}O_2$ kinetics

Table 1 presents the group mean \pm SD parameter estimates for $\dot{V}O_2$ kinetics with their corresponding profiles in a representative participant shown in Figure 1. There was no significant difference in the $\tau\dot{V}O_{2p}$ between U \rightarrow M and U \rightarrow VH ($P = 0.31$, $ES = 0.4$); however, the $\tau\dot{V}O_{2p}$ was slower during M \rightarrow VH compared to other conditions ($P < 0.001$, $ES > 1.2$). There was a significant main effect for step cycling on the G_p , which, relative to U \rightarrow M, decreased during U \rightarrow VH ($P = 0.01$, $ES = 0.7$); however, there were no significant differences during M \rightarrow VH compared to other conditions ($P > 0.2$). The A_{Sc} decreased during M \rightarrow VH compared to U \rightarrow VH ($P = 0.01$, $ES = 0.8$) with this difference removed when normalizing A_{Sc} relative to the total $\Delta\dot{V}O_2$ above baseline pedaling (U \rightarrow VH: 14 ± 6 vs. M \rightarrow VH: 13 ± 7 %, $P = 0.37$). Relative to U \rightarrow M, the G_{tot} was greater during U \rightarrow VH ($P = 0.045$, $ES = 0.6$) and M \rightarrow VH ($P = 0.03$, $ES = 0.9$).

NIRS-derived variables

Table 2 presents the group mean \pm SD parameter estimates for NIRS-derived deoxy[Hb+Mb] and TOI kinetics with their corresponding profiles in a representative participant shown in Figures 2 and 3, respectively. Relative to U \rightarrow M, the $\Delta\text{deoxy[Hb+Mb]}-TD$ following exercise onset decreased in the other conditions ($P < 0.001$, $ES > 1.9$) with a further reduction during M \rightarrow VH compared to U \rightarrow VH ($P = 0.03$, $ES = 0.8$). There were no significant differences ($P > 0.40$) between U \rightarrow M and U \rightarrow VH in the $\tau\Delta\text{deoxy[Hb+Mb]}$ or τTOI ; however, both were slowed during M \rightarrow VH compared to

other conditions ($P < 0.03$, $ES > 1.2$). Accordingly, the overall MRT (i.e. $TD + \tau$) of muscle deoxygenation kinetics was faster during $U \rightarrow VH$ compared to $U \rightarrow M$ and $M \rightarrow VH$ ($P < 0.001$, $ES > 1.2$).

Matching of deoxy[Hb+Mb] to $\dot{V}O_2$

Comparison of the group mean \pm SD kinetic parameters for $\dot{V}O_2$ and muscle deoxygenation are presented in Figure 4. There were no significant differences between the $\tau\dot{V}O_{2p}$ and muscle deoxygenation kinetics during $U \rightarrow M$ ($P > 0.15$), whereas, the MRT of $\Delta\text{deoxy[Hb+Mb]}$ and TOI was speeded relative to the $\tau\dot{V}O_{2p}$ during $U \rightarrow VH$ and $M \rightarrow VH$ ($P < 0.001$). The difference between the $\tau\dot{V}O_{2p}$ and $MRT-\Delta\text{deoxy[Hb+Mb]}$ increased by a large effect size during $M \rightarrow VH$ compared to $U \rightarrow VH$ (18 ± 15 vs. 9 ± 7 s, $P = 0.07$, $ES = 0.8$, Figure 4C), with a significantly greater difference between the $\tau\dot{V}O_{2p}$ and $MRT\text{-TOI}$ during work-to-work exercise (23 ± 15 vs. 12 ± 7 s, respectively, $P = 0.014$, $ES = 1.0$, Figure 4D). During $U \rightarrow M$, the normalized $\Delta\text{deoxy[Hb+Mb]}/\Delta\dot{V}O_2$ overshoot area yielded non-normally distributed data; therefore, were not reported. As shown in Figure 5, the overshoot area above unity in the normalized $\Delta\text{deoxy[Hb+Mb]}/\Delta\dot{V}O_2$ ratio was significantly greater during the on-transition of $M \rightarrow VH$ compared to $U \rightarrow VH$ exercise (17.3 ± 13.2 vs. 8.5 ± 7.0 %/s, $P = 0.01$, $ES = 0.9$, respectively).

Relationship between $\dot{V}O_2$ and deoxy[Hb+Mb] kinetics

The reduction of the G_p correlated positively with the speeding of the $MRT-\Delta\text{deoxy[Hb+Mb]}$ during $U \rightarrow VH$ compared to $U \rightarrow M$ ($r = 0.67$; $P = 0.005$). During $M \rightarrow VH$, there was no significant relationship ($P > 0.5$) between the slowing of the $\tau\dot{V}O_{2p}$ with

alterations in the $\tau\Delta\text{deoxy[Hb+Mb]}$ compared to U→M or U→VH ($r = 0.15$ and -0.06 , respectively).

Relationship between phase II $\dot{V}\text{O}_2$ with chronological age and baseline $\dot{V}\text{O}_2$

There was no significant relationship between the $\tau\dot{V}\text{O}_{2p}$ with chronological age during U→M ($r = 0.40$, $P = 0.09$); however, both variables correlated positively during U→VH ($r = 0.48$, $P = 0.04$) with a stronger relationship observed during M→VH ($r = 0.78$, $P < 0.001$). An increased (Δ) $\tau\dot{V}\text{O}_{2p}$ and ΔG_p during M→VH relative to U→M and U→VH correlated positively with chronological age ($P < 0.01$, Figure 6 A-D). During M→VH, the baseline $\dot{V}\text{O}_2$ in l/min correlated positively with the ΔG_p relative to U→M ($r = 0.59$, $P = 0.008$) and U→VH ($r = 0.71$, $P = 0.001$); however, there was no significant relationship with the $\Delta\tau\dot{V}\text{O}_{2p}$ relative to other conditions ($r = 0.44$ and 0.39 , $P = 0.07$ and 0.11 , respectively).

DISCUSSION

This study combined simultaneous measurements of $\dot{V}\text{O}_2$ and NIRS-derived muscle deoxygenation kinetics to investigate the relationship between dynamic (non)linearity of the $\tau\dot{V}\text{O}_{2p}$ and G_p with alterations in skeletal muscle O_2 delivery/utilization during step exercise in 11 to 15 y olds. In line with our study hypothesis, relative to U→M cycling, an invariant $\tau\dot{V}\text{O}_{2p}$ during U→VH was accompanied by a faster $MRT\text{-}\Delta\text{deoxy[Hb+Mb]}$ and $MRT\text{-TOI}$, suggesting that an increased rate of fractional O_2 extraction mitigated a decreased $\dot{Q}\text{O}_2/\dot{V}\text{O}_2$ response during supra-LT transitions initiated from unloaded pedaling. However, during U→VH compared U→M, the G_p decreased suggesting that

this parameter may be limited by decreased microvascular O_2 delivery in boys. Conversely, relative to $U \rightarrow VH$ cycling, $M \rightarrow VH$ decreased the rate of fractional O_2 extraction (i.e. increased MRT of $\Delta\text{deoxy[Hb+Mb]}$ and TOI kinetics) in a manner that was disproportionately less than the slowing of the $\tau \dot{V}O_{2p}$, thereby, eliciting a greater $\Delta\text{deoxy[Hb+Mb]}/\Delta\dot{V}O_2$ “overshoot” in the transition from a raised baseline work rate. Finally, relative to $U \rightarrow M$ and $U \rightarrow VH$, an increased $\tau \dot{V}O_{2p}$ and G_p during $M \rightarrow VH$ correlated positively with boys’ chronological age. These findings lend support to the notion that developmental effects on $\dot{V}O_2$ kinetics might be linked to the recruitment of higher-order (i.e. type II) muscle fibers with slower microvascular blood flow dynamics and poorer efficiency in older youth.

Comparison of $\dot{V}O_2$ and muscle deoxygenation kinetics between $U \rightarrow M$ and $U \rightarrow VH$

In the present study, we observed no significant differences in the $\tau \dot{V}O_{2p}$ between $U \rightarrow M$ and $U \rightarrow VH$; however, the G_p decreased during $U \rightarrow VH$ reflecting both linear and non-linear control features of $\dot{V}O_2$ kinetics following the onset of step cycling elicited from unloaded pedaling in 11 to 15 y olds. An invariant $\tau \dot{V}O_{2p}$ during different intensities of step exercise is consistent with previous reports in youth (Hebestreit *et al.*, 1998; Williams *et al.*, 2001; Lai *et al.*, 2008); however, these studies employed relatively low sample sizes (i.e. $n = 8$), or, in the case of Hebestreit *et al.* (1998) the work rate was arbitrarily normalized as a fraction of peak $\dot{V}O_2$ in children. Therefore, our findings extend those previously reported by revealing an invariant $\tau \dot{V}O_{2p}$ relative to work rate using procedures for resolving the kinetic parameters within carefully prescribed

intensity domains among a larger youth cohort (i.e. $n = 19$), hence, reducing the potential for type II statistical error.

Following the onset of $U \rightarrow M$ and $U \rightarrow VH$, there was a pronounced TD before muscle deoxy[Hb+Mb] increased, suggesting that the hyperemic effect of skeletal muscle contractions sufficiently matched the requirement for O_2 utilization within active regions of vastus lateralis muscle. However, during $U \rightarrow VH$ compared to $U \rightarrow M$, the MRT - Δ deoxy[Hb+Mb] and MRT -TOI were reduced (i.e. decreased $TD + \tau$) by a large effect size, hence, suggesting that an increased rate of fractional O_2 extraction was required to maintain an invariant $\tau \dot{V}O_{2p}$ between both conditions. Therefore, these findings, in line with “priming” exercise studies in children (Barker *et al.*, 2010; Barker *et al.*, 2014), support the notion that the $\tau \dot{V}O_{2p}$ is principally limited by intracellular metabolic factors rather than the dynamic relationship between $\dot{Q}O_2$ -to- $\dot{V}O_2$ during supra-LT transitions in youth.

In the present study, we did observe a significant association between a decreased G_p with the relative speeding of the MRT - Δ deoxy[Hb+Mb] following the onset of $U \rightarrow VH$ compared to $U \rightarrow M$. It has been reported that the τ of deoxy[Hb+Mb] kinetics coheres with that observed for the reduction in microvascular O_2 partial pressure ($P_{mv}O_2$) following the onset of skeletal muscle contractions (Koga *et al.*, 2012). Accordingly, it is conceivable that those participants evincing a greater $\dot{Q}O_2$ -to- $\dot{V}O_2$ mismatch (i.e. faster MRT - Δ deoxy[Hb+Mb]) might have accelerated the fall in $P_{mv}O_2$ such that the $\dot{V}O_2$ increment per unit of work rate was limited consequent to a decreased O_2 flux between the capillary and muscle. Therefore, in youth, our findings are consistent with the notion

that the G_p might be sensitive to a decreased $\dot{Q}_{O_2}/\dot{V}O_2$ response as previously reported in adults (Koga *et al.*, 1999; Jones *et al.*, 2006).

Comparison of $\dot{V}O_2$ and muscle deoxygenation kinetics during M→VH relative to other step conditions

Whereas U→VH sped muscle deoxy[Hb+Mb] kinetics, to maintain a constant $\tau\dot{V}O_{2p}$ compared to U→M, there was a concomitant slowing of the $\tau\dot{V}O_{2p}$, τTOI and $\tau\Delta\text{deoxy[Hb+Mb]}$ during M→VH compared to other step conditions. Whilst these findings during M→VH are consistent with a decreased rate of O_2 extraction, it is important to consider that U→VH and M→VH sped the MRT of $\Delta\text{deoxy[Hb+Mb]}$ and TOI responses relative to the $\tau\dot{V}O_{2p}$ with the difference between these signals increased following the onset of work-to-work transitions (Figure 4). In other words, the slowing of muscle deoxygenation did not match proportionally the slowing of the $\tau\dot{V}O_{2p}$, thereby, increasing the normalized $\Delta\text{deoxy[Hb+Mb]}/\Delta\dot{V}O_2$ overshoot area above unity within the initial few minutes of M→VH compared to U→VH (Figure 5). Collectively, these responses during M→VH are consistent with an increased proportional reliance on fractional O_2 extraction; hence, our results suggest for the first time in youth that slower phase II $\dot{V}O_2$ kinetics coincided with a slower rate of adjustment in \dot{Q}_{O_2} -to- $\dot{V}O_2$ in the transition from a raised baseline work rate.

In boys, it had been previously suggested that eliciting step transitions from a raised level of electromyogram activity increased proportionally the recruitment of type II muscle fibers for power production (Breese *et al.*, 2012). This supposition was based on an orderly 'size' principle of motor unit recruitment (Henneman & Mendell, 1981), which,

in adults, has received support with previous studies reporting a progressive reduction in the glycogen content within type I followed by type IIa and IIx muscle fibers from low to high force requirements (Essen, 1978; Green, 1978; Krstrup *et al.*, 2004). ‘Higher-order’ type II muscle fibers have been reported to possess slower microvascular O₂ delivery (i.e. decreased P_{mvO_2} across the on-exercise transition) (Behnke *et al.*, 2003) and slower $\dot{V}O_2$ kinetics *in vitro* compared with ‘lower-order’ type I muscle fibers (Crow & Kushmerick, 1982). Therefore, during M→VH, it is conceivable that the $\dot{V}O_2$ and deoxy[Hb+Mb] profiles (and their kinetic relationship) reflected the intrinsic properties of a population of skeletal muscle fibers positioned higher in the recruitment hierarchy in boys.

There was a significant main effect for step cycling on the G_{tot} , which, relative to U→M, was greater during U→VH and M→VH consequent to the development of the A_{Sc} in these conditions. However, relative to U→VH, the A_{Sc} decreased by ~ 50% during M→VH such that $\dot{V}O_2$ kinetics reverted toward a mono-exponential profile. There is evidence to suggest that the development of the A_{Sc} is related in some manner to the recruitment profile and metabolic features of type II muscle fibers with slower $\dot{V}O_2$ kinetics and poorer efficiency [i.e. increased ATP/force output ratio (Crow & Kushmerick, 1982)] compared with type I muscle fibers (see Jones *et al.*, 2011 for review). Therefore, in adults, an explanation for a smaller A_{Sc} has considered the earlier (rather than latent) expression upon the pulmonary $\dot{V}O_2$ signal of higher-order (i.e. type II) muscle fibers when supra-LT transitions are initiated from an elevated work rate (Wilkerson & Jones, 2007; DiMenna *et al.*, 2008). However, this proposal predicts that the G_p would have

been higher during M→VH relative to other conditions, which, in boys, was not present with this effect associated with chronological age (Figure 6).

Relationship between $\dot{V}O_2$ kinetics with chronological age

A novel finding was that the $\tau\dot{V}O_{2p}$ and chronological age, whilst not significantly associated during U→M, were both positively correlated during U→VH with this relationship strengthened by an increased pre-transition work rate. In other words, M→VH exercise slowed by a greater extent the $\tau\dot{V}O_{2p}$ and increased the G_p within the age range between 11 to 15 y (Figure 6). It would have been expected that U→M transitions predominantly recruited a population of type I muscle fibers (Krustrup *et al.*, 2004) with the mean $\tau\dot{V}O_{2p}$ in this condition in boys (i.e. ~ 23 s) less likely to be limited by muscle O_2 delivery based on a previous study in adults (Murias *et al.*, 2011). Conversely, a previous investigation has reported a slower $\tau\dot{V}O_{2p}$ alongside slower limb blood flow dynamics following the onset of work-to-work exercise in adults (MacPhee *et al.*, 2005) with further evidence in support of a decline in the maximal rate of O_2 transport between the ages of 12 to 17 y (Koch, 1984) and in the proportion of type I muscle fibers within the vastus lateralis between the ages of 5 to 20 y (Lexell *et al.*, 1992). Therefore, we propose indirectly that an age-related slowing of the $\tau\dot{V}O_{2p}$ during M→VH might have reflected differences in muscle perfusion and the distribution of O_2 in conjunction with alterations in muscle fiber recruitment in older youth.

Alternatively, it is important to consider that larger (older) boys produced higher cycling power outputs corresponding to the GET and at task failure during the initial ramp incremental test. Therefore, during M→VH, it would have been expected that

baseline pedaling equivalent to 90% GET recruited a larger muscle mass resulting in a greater pre-transition $\dot{V}O_2$ compared to smaller (younger) children. In this regard, it has been reported that the $\tau\dot{V}O_{2p}$ and G_p increased linearly at progressively higher baseline power outputs (hence $\dot{V}O_2$) in adults (Keir *et al.*, 2016), providing an additional explanation for the relationships presented in Figure 6. However, we reported no significant association between baseline $\dot{V}O_2$ in l/min during M→VH with the $\Delta\tau\dot{V}O_{2p}$ relative to U→M and U→VH exercise.

Assuming that U→M immediately followed by M→VH evoked an orderly recruitment of motor units, the relationships presented in Figure 6 lend support to the notion that work-to-work cycling revealed a greater disparity in the τ and G values between higher- relative to lower-order muscle fibers with increased chronological age (Figure 7). Accordingly, if the measured $\dot{V}O_2$ profile during U→VH reflected the summed response of muscle fiber pools recruited separately during U→M and M→VH (Wilkerson & Jones, 2007), then those positioned higher in the recruitment hierarchy (i.e. type II) would be expected to elicit a net slowing of pulmonary $\dot{V}O_2$ during the on-transition of exercise and/or extend the A_{Sc} in older children. This $\dot{V}O_2$ response is characteristic of that previously observed longitudinally in youth (Fawkner & Armstrong, 2004; Breese *et al.*, 2010); therefore, our findings shed potential novel insight into the physiological factors responsible for modulating $\dot{V}O_2$ kinetics between the ages of 11 to 15 y.

Limitations

It is recognized that there exist limitations with CW-NIRS assuming constant tissue optical properties (i.e. path length, absorption and scattering coefficients), which, has

been reported to confound interpretation of deoxy[Hb+Mb] data (see Barstow *et al.*,
 2019 for a recent review). Moreover, we also recognize that the absorbance spectra of
 Hb and Mb overlap within the NIR range; therefore, the relative (%) contribution from
 each chromophore to the NIRS-derived signal is uncertain (Masuda *et al.*, 2010; Davis &
 Barstow, 2013). Additionally, we left shifted the normalized $\dot{V}O_2$ by 15 s to account for
 the estimated phase I duration in children (Springer *et al.*, 1991; Hebestreit *et al.*, 1998),
 thereby, time aligning the start of phase II $\dot{V}O_2$ to the onset of exercise, which, has been
 reported to coincide with muscle $\dot{V}O_2$ within 10% (Barstow *et al.*, 1994). Therefore, the
 extent to which inter- and intra-participant differences in the circulatory muscle-to-lung
 transit time influenced the $\Delta\text{deoxy[Hb+Mb]}/\Delta\dot{V}O_2$ overshoot is unclear. It should also be
 cautioned that the pulmonary $\dot{V}O_2$ amplitude during exercise includes minor
 contributions from cardiorespiratory support processes (Poole *et al.*, 1991), which, has
 the potential to influence its ratio when expressed relative to the adjustment in
 deoxy[Hb+Mb] kinetics. Therefore, in the present study, we stress that precedence be
 given to interpreting the TD and τ of muscle deoxygenation with these preliminary
 kinetic data supported by the $\Delta\text{deoxy[Hb+Mb]}/\Delta\dot{V}O_2$ ratio to infer the dynamic
 (mis)matching between O_2 delivery/utilization. Finally, it should be noted that baseline
 pedaling during M→VH involved simultaneously raising pre-transition $\dot{V}O_2$ with work
 rate, which, when both are dissociated, has the potential to influence the $\tau\dot{V}O_{2p}$ and G_p
 via independent mechanisms (DiMenna *et al.*, 2010a; Bowen *et al.*, 2011; Wust *et al.*,
 2014). Therefore, in the present study, whether an increased baseline work rate *per se*
 altered phase II $\dot{V}O_2$ kinetics cannot be established.

Conclusions

This study in 11 to 15 y olds reported dynamic non-linearity of the phase II $\dot{V}O_2$ kinetic parameters, with respect to a decreased G_p during U→VH compared to U→M, whereas, a slower $\tau\dot{V}O_{2p}$ was dependent on an increased pre-transition work rate in youth. Furthermore, whilst “work-to-work” cycling slowed the τ of muscle deoxygenation, when expressed relative to the adjustment in $\dot{V}O_2$ kinetics, the ratio between both of these signals increased, suggesting a greater proportional speeding of fractional O_2 extraction; hence, the slower $\tau\dot{V}O_{2p}$ during M→VH was consequent to a slowing of microvascular blood flow relative to O_2 utilization. Finally, an increased $\tau\dot{V}O_{2p}$ and G_p during the transition from a raised baseline work rate correlated positively with chronological age. These novel findings further our understanding of the physiological factors modulating the $\dot{V}O_2$ kinetic response, and, thereby, oxidative metabolism, and their association with chronological age in healthy youth.

REFERENCES

- Babcock MA, Paterson DH, Cunningham DA & Dickinson JR (1994). Exercise on-transient gas exchange kinetics are slowed as a function of age. *Med Sci Sports Exerc* **26**, 440-446.
- Barker AR, Jones AM & Armstrong N (2010). The influence of priming exercise on oxygen uptake, cardiac output, and muscle oxygenation kinetics during very heavy-intensity exercise in 9- to 13-yr-old boys. *J Appl Physiol* **109**, 491-500.
- Barker AR, Trebilcock E, Breese B, Jones AM & Armstrong N (2014). The effect of priming exercise on O₂ uptake kinetics, muscle O₂ delivery and utilization, muscle activity, and exercise tolerance in boys. *Appl Physiol Nutr Metab* **39**, 308-317.
- Barker AR, Welsman JR, Fulford J, Welford D, Williams CA & Armstrong N (2008). Muscle phosphocreatine and pulmonary oxygen uptake kinetics in children at the onset and offset of moderate intensity exercise. *Eur J Appl Physiol* **102**, 727-738.
- Barker AR, Williams CA, Jones AM & Armstrong N (2011). Establishing maximal oxygen uptake in young people during a ramp cycle test to exhaustion. *Br J Sports Med* **45**, 498-503.
- Barstow TJ (2019). Understanding near infrared spectroscopy and its application to skeletal muscle research. *J Appl Physiol* **126**, 1360-1376.
- Barstow TJ, Buchthal S, Zanconato S & Cooper DM (1994). Muscle energetics and pulmonary oxygen uptake kinetics during moderate exercise. *J Appl Physiol* **77**, 1742-1749.
- Barstow TJ, Jones AM, Nguyen PH & Casaburi R (1996). Influence of muscle fiber type and pedal frequency on oxygen uptake kinetics of heavy exercise. *J Appl Physiol* **81**, 1642-1650.
- Beaver WL, Wasserman K & Whipp BJ (1986). A new method for detecting anaerobic threshold by gas exchange. *J Appl Physiol* **60**, 2020-2027.
- Behnke BJ, McDonough P, Padilla DJ, Musch TI & Poole DC (2003). Oxygen exchange profile in rat muscles of contrasting fibre types. *J Physiol* **549**, 597-605.

587 Benson AP, Grassi B & Rossiter HB (2013). A validated model of oxygen uptake and
 588 circulatory dynamic interactions at exercise onset in humans. *J Appl Physiol* **115**, 743-
 589 755.

590

591 Bowen TS, Murgatroyd SR, Cannon DT, Cuff TJ, Lainey AF, Marjerrison AD, Spencer MD,
 592 Benson AP, Paterson DH, Kowalchuk JM & Rossiter HB (2011). A raised metabolic rate
 593 slows pulmonary O₂ uptake kinetics on transition to moderate-intensity exercise in
 594 humans independently of work rate. *Exp Physiol* **96**, 1049-1061.

595

596 Breese BC, Barker AR, Armstrong N, Jones AM & Williams CA (2012). The effect of
 597 baseline metabolic rate on pulmonary O₂ uptake kinetics during very heavy intensity
 598 exercise in boys and men. *Respir Physiol Neurobiol* **180**, 223-229.

599

600 Breese BC, Williams CA, Barker AR, Welsman JR, Fawkner SG & Armstrong N (2010).
 601 Longitudinal change in the oxygen uptake kinetic response to heavy-intensity exercise
 602 in 14- to 16-years-old boys. *Pediatr Exerc Sci* **22**, 314-325.

603

604 Brittain CJ, Rossiter HB, Kowalchuk JM & Whipp BJ (2001). Effect of prior metabolic rate
 605 on the kinetics of oxygen uptake during moderate-intensity exercise. *Eur J Appl Physiol*
 606 **86**, 125-134.

607

608 Crow MT & Kushmerick MJ (1982). Chemical energetics of slow- and fast-twitch muscles
 609 of the mouse. *J Gen Physiol* **79**, 147-166.

610

611 Davis ML & Barstow TJ (2013). Estimated contribution of hemoglobin and myoglobin to
 612 near infrared spectroscopy. *Respir Physiol Neurobiol* **186**, 180-187.

613

614 DeLorey DS, Kowalchuk JM & Paterson DH (2003). Relationship between pulmonary O₂
 615 uptake kinetics and muscle deoxygenation during moderate-intensity exercise. *J Appl*
 616 *Physiol* **95**, 113-120.

617

618 DeLorey DS, Kowalchuk JM & Paterson DH (2005). Adaptation of pulmonary O₂ uptake
 619 kinetics and muscle deoxygenation at the onset of heavy-intensity exercise in young and
 620 older adults. *J Appl Physiol* **98**, 1697-1704.

621

622 DiMenna FJ, Bailey SJ, Vanhatalo A, Chidnok W & Jones AM (2010a). Elevated baseline
 623 $\dot{V}O_2$ per se does not slow O₂ uptake kinetics during work-to-work exercise transitions. *J*
 624 *Appl Physiol* **109**, 1148-1154.

625
626 DiMenna FJ, Fulford J, Bailey SJ, Vanhatalo A, Wilkerson DP & Jones AM (2010b).
627 Influence of priming exercise on muscle [PCr] and pulmonary O₂ uptake dynamics during
628 'work-to-work' knee-extension exercise. *Respir Physiol Neurobiol* **172**, 15-23.

629
630 DiMenna FJ, Wilkerson DP, Burnley M & Jones AM (2008). Influence of priming exercise
631 on pulmonary O₂ uptake kinetics during transitions to high-intensity exercise from an
632 elevated baseline. *J Appl Physiol* **105**, 538-546.

633
634 Essen B (1978). Glycogen depletion of different fibre types in human skeletal muscle
635 during intermittent and continuous exercise. *Acta Physiol Scand* **103**, 446-455.

636
637 Fawcner SG & Armstrong N (2004). Longitudinal changes in the kinetic response to
638 heavy-intensity exercise in children. *J Appl Physiol* **97**, 460-466.

639
640 Fujihara Y, Hildebrandt J & Hildebrandt JR (1973a). Cardiorespiratory transients in
641 exercising man. II. Linear models. *J Appl Physiol* **35**, 68-76.

642
643 Fujihara Y, Hildebrandt JR & Hildebrandt J (1973b). Cardiorespiratory transients in
644 exercising man. I. Tests of superposition. *J Appl Physiol* **35**, 58-67.

645
646 Grassi B, Pogliaghi S, Rampichini S, Quaresima V, Ferrari M, Marconi C & Cerretelli P
647 (2003). Muscle oxygenation and pulmonary gas exchange kinetics during cycling exercise
648 on-transitions in humans. *J Appl Physiol* **95**, 149-158.

649
650 Grassi B, Poole DC, Richardson RS, Knight DR, Erickson BK & Wagner PD (1996). Muscle
651 O₂ uptake kinetics in humans: implications for metabolic control. *J Appl Physiol* **80**, 988-
652 998.

653
654 Green HJ (1978). Glycogen depletion patterns during continuous and intermittent ice
655 skating. *Med Sci Sports* **10**, 183-187.

656
657 Hebestreit H, Kriemler S, Hughson RL & Bar-Or O (1998). Kinetics of oxygen uptake at
658 the onset of exercise in boys and men. *J Appl Physiol* **85**, 1833-1841.

659
660 Henneman E & Mendell LM (1981). Functional Organisation of a Motoneuron Pool and
661 its Inputs. In: *Handbook of Physiology I, Vol II, Part 1* edited by Brooks VB, pp. 423-507.
662 American Physiological Society, Bethesda, U.S.

663
 664 Hughson RL & Morrissey M (1982). Delayed kinetics of respiratory gas exchange in the
 665 transition from prior exercise. *J Appl Physiol* **52**, 921-929.

666
 667 Hughson RL, Tschakovsky ME & Houston ME (2001). Regulation of oxygen consumption
 668 at the onset of exercise. *Exerc Sport Sci Rev* **29**, 129-133.

669
 670 Jones AM, Berger NJ, Wilkerson DP & Roberts CL (2006). Effects of "priming" exercise on
 671 pulmonary O₂ uptake and muscle deoxygenation kinetics during heavy-intensity cycle
 672 exercise in the supine and upright positions. *J Appl Physiol* **101**, 1432-1441.

673
 674 Jones AM, Grassi B, Christensen PM, Krstrup P, Bangsbo J & Poole DC (2011). Slow
 675 component of $\dot{V}O_2$ kinetics: mechanistic bases and practical applications. *Med Sci Sports*
 676 *Exerc* **43**, 2046-2062.

677
 678 Jones AM, Wilkerson DP & Fulford J (2008). Muscle [phosphocreatine] dynamics
 679 following the onset of exercise in humans: the influence of baseline work-rate. *J Physiol*
 680 **586**, 889-898.

681
 682 Keir DA, Robertson TC, Benson AP, Rossiter HB & Kowalchuk JM (2016). The influence of
 683 metabolic and circulatory heterogeneity on the expression of pulmonary oxygen uptake
 684 kinetics in humans. *Exp Physiol* **101**, 176-192.

685
 686 Koch G (1984). Maximal Oxygen Transport Capacity in Adolescents Aged 12 to 17 Years.
 687 Effect of Growth Combined with Intensive Physical Training. In: *Current Topics in Sports*
 688 *Medicine* edited by Bachl N, Prokop L & Suckert R, pp. 479-497. Urban and
 689 Schwarzenberg, Wien.

690
 691 Koga S, Kano Y, Barstow TJ, Ferreira LF, Ohmae E, Sudo M & Poole DC (2012). Kinetics of
 692 muscle deoxygenation and microvascular PO₂ during contractions in rat: comparison of
 693 optical spectroscopy and phosphorescence-quenching techniques. *J Appl Physiol* **112**,
 694 26-32.

695
 696 Koga S, Poole DC, Shiojiri T, Kondo N, Fukuba Y, Miura A & Barstow TJ (2005).
 697 Comparison of oxygen uptake kinetics during knee extension and cycle exercise. *Am J*
 698 *Physiol Regul Integr Comp Physiol* **288**, R212-220.

699
 700 Koga S, Shiojiri T, Shibasaki M, Kondo N, Fukuba Y & Barstow TJ (1999). Kinetics of oxygen
 701 uptake during supine and upright heavy exercise. *J Appl Physiol* **87**, 253-260.

702
703 Koppo K, Bouckaert J & Jones AM (2004). Effects of training status and exercise intensity
704 on phase II $\dot{V}O_2$ kinetics. *Med Sci Sports Exerc* **36**, 225-232.

705
706 Krstrup P, Jones AM, Wilkerson DP, Calbet JA & Bangsbo J (2009). Muscular and
707 pulmonary O_2 uptake kinetics during moderate- and high-intensity sub-maximal knee-
708 extensor exercise in humans. *J Physiol* **587**, 1843-1856.

709
710 Krstrup P, Soderlund K, Mohr M & Bangsbo J (2004). The slow component of oxygen
711 uptake during intense, sub-maximal exercise in man is associated with additional fibre
712 recruitment. *Pflugers Arch* **447**, 855-866.

713
714 Lai N, Nasca MM, Silva MA, Silva FT, Whipp BJ & Cabrera ME (2008). Influence of exercise
715 intensity on pulmonary oxygen uptake kinetics at the onset of exercise and recovery in
716 male adolescents. *Appl Physiol Nutr Metab* **33**, 107-117.

717
718 Lexell J, Sjostrom M, Nordlund AS & Taylor CC (1992). Growth and development of
719 human muscle: a quantitative morphological study of whole vastus lateralis from
720 childhood to adult age. *Muscle Nerve* **15**, 404-409.

721
722 MacPhee SL, Shoemaker JK, Paterson DH & Kowalchuk JM (2005). Kinetics of O_2 uptake,
723 leg blood flow, and muscle deoxygenation are slowed in the upper compared with lower
724 region of the moderate-intensity exercise domain. *J Appl Physiol* **99**, 1822-1834.

725
726 Masuda K, Takakura H, Furuichi Y, Iwase S & Jue T (2010). NIRS measurement of O_2
727 dynamics in contracting blood and buffer perfused hindlimb muscle. *Adv Exp Med Biol*
728 **662**, 323-328.

729
730 McNarry MA (2019). Oxygen uptake kinetics in youth: characteristics, interpretation,
731 and application. *Pediatr Exerc Sci* **31**, 175-183.

732
733 McNarry MA, Kingsley MI & Lewis MJ (2012). Influence of exercise intensity on
734 pulmonary oxygen uptake kinetics in young and late middle-aged adults. *Am J Physiol*
735 *Regul Integr Comp Physiol* **303**, R791-798.

736
737 Moore SA, McKay HA, Macdonald H, Nettlefold L, Baxter-Jones AD, Cameron N &
738 Brasher PM (2015). Enhancing a somatic maturity prediction model. *Med Sci Sports Exerc*
739 **47**, 1755-1764.

740

741 Murias JM, Kowalchuk JM & Paterson DH (2010). Speeding of $\dot{V}O_2$ kinetics with

742 endurance training in old and young men is associated with improved matching of local

743 O_2 delivery to muscle O_2 utilization. *J Appl Physiol* **108**, 913-922.

744

745 Murias JM, Spencer MD, Kowalchuk JM & Paterson DH (2011). Muscle deoxygenation to

746 $\dot{V}O_2$ relationship differs in young subjects with varying $\tau\dot{V}O_2$. *Eur J Appl Physiol* **111**,

747 3107-3118.

748

749 Murias JM, Spencer MD & Paterson DH (2014). The critical role of O_2 provision in the

750 dynamic adjustment of oxidative phosphorylation. *Exerc Sport Sci Rev* **42**, 4-11.

751

752 Ozyener F, Rossiter HB, Ward SA & Whipp BJ (2001). Influence of exercise intensity on

753 the on- and off-transient kinetics of pulmonary oxygen uptake in humans. *J Physiol* **533**,

754 891-902.

755

756 Paterson DH & Whipp BJ (1991). Asymmetries of oxygen uptake transients at the on-

757 and offset of heavy exercise in humans. *J Physiol* **443**, 575-586.

758

759 Poole DC, Schaffartzik W, Knight DR, Derion T, Kennedy B, Guy HJ, Prediletto R & Wagner

760 PD (1991). Contribution of exercising legs to the slow component of oxygen uptake

761 kinetics in humans. *J Appl Physiol* **71**, 1245-1260.

762

763 Pringle JS, Doust JH, Carter H, Tolfrey K, Campbell IT, Sakkas GK & Jones AM (2003).

764 Oxygen uptake kinetics during moderate, heavy and severe intensity "submaximal"

765 exercise in humans: the influence of muscle fibre type and capillarisation. *Eur J Appl*

766 *Physiol* **89**, 289-300.

767

768 Quaresima V & Ferrari M (2009). Muscle oxygenation by near-infrared-based tissue

769 oximeters. *J Appl Physiol (1985)* **107**, 371; author reply 372-373.

770

771 Ratel S, Tonson A, Le Fur Y, Cozzone P & Bendahan D (2008). Comparative analysis of

772 skeletal muscle oxidative capacity in children and adults: a ^{31}P -MRS study. *Appl Physiol*

773 *Nutr Metab* **33**, 720-727.

774

775 Rossiter HB, Ward SA, Doyle VL, Howe FA, Griffiths JR & Whipp BJ (1999). Inferences

776 from pulmonary O_2 uptake with respect to intramuscular [phosphocreatine] kinetics

777 during moderate exercise in humans. *J Physiol* **518 (Pt 3)**, 921-932.

778
 779 Rossiter HB, Ward SA, Kowalchuk JM, Howe FA, Griffiths JR & Whipp BJ (2001). Effects
 780 of prior exercise on oxygen uptake and phosphocreatine kinetics during high-intensity
 781 knee-extension exercise in humans. *J Physiol* **537**, 291-303.

782
 783 Sansum KM, Weston ME, Bond B, Cockcroft EJ, O'Connor A, Tomlinson OW, Williams CA
 784 & Barker AR (2019). Validity of the supramaximal test to verify maximal oxygen uptake
 785 in children and adolescents. *Pediatr Exerc Sci* **31**, 213-222.

786
 787 Spencer MD, Murias JM, Grey TM & Paterson DH (2012). Regulation of $\dot{V}O_2$ kinetics by
 788 O_2 delivery: insights from acute hypoxia and heavy-intensity priming exercise in young
 789 men. *J Appl Physiol* **112**, 1023-1032.

790
 791 Springer C, Barstow TJ, Wasserman K & Cooper DM (1991). Oxygen uptake and heart
 792 rate responses during hypoxic exercise in children and adults. *Med Sci Sports Exerc* **23**,
 793 71-79.

794
 795 Tonson A, Ratel S, Le Fur Y, Vilmen C, Cozzone PJ & Bendahan D (2010). Muscle
 796 energetics changes throughout maturation: a quantitative ^{31}P -MRS analysis. *J Appl*
 797 *Physiol* **109**, 1769-1778.

798
 799 Whipp BJ, Davis JA, Torres F & Wasserman K (1981). A test to determine parameters of
 800 aerobic function during exercise. *J Appl Physiol* **50**, 217-221.

801
 802 Wilkerson DP & Jones AM (2006). Influence of initial metabolic rate on pulmonary O_2
 803 uptake on-kinetics during severe intensity exercise. *Respir Physiol Neurobiol* **152**, 204-
 804 219.

805
 806 Wilkerson DP & Jones AM (2007). Effects of baseline metabolic rate on pulmonary O_2
 807 uptake on-kinetics during heavy-intensity exercise in humans. *Respir Physiol Neurobiol*
 808 **156**, 203-211.

809
 810 Wilkerson DP, Koppo K, Barstow TJ & Jones AM (2004). Effect of work rate on the
 811 functional 'gain' of phase II pulmonary O_2 uptake response to exercise. *Respir Physiol*
 812 *Neurobiol* **142**, 211-223.

813
 814 Williams CA, Carter H, Jones AM & Doust JH (2001). Oxygen uptake kinetics during
 815 treadmill running in boys and men. *J Appl Physiol* **90**, 1700-1706.

Wust RC, McDonald JR, Sun Y, Ferguson BS, Rogatzki MJ, Spires J, Kowalchuk JM, Gladden LB & Rossiter HB (2014). Slowed muscle oxygen uptake kinetics with raised metabolism are not dependent on blood flow or recruitment dynamics. *J Physiol* **592**, 1857-1871.

ADDITIONAL INFORMATION

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AUTHOR CONTRIBUTIONS

Conception or design of the work: B.C.B. and C.A.W. Acquisition, analysis or interpretation of data for the work and revising it critically for important intellectual content: all authors. All authors approved the final version of the manuscript and agree to be accountable for all aspects of the work in ensuring that questions related to the accuracy or integrity of any part of the work are appropriately investigated and resolved. All persons designated as authors qualify for authorship, and all those who qualify for authorship are listed.

TABLES

Table 1. Amplitude and kinetics of pulmonary oxygen uptake ($\dot{V}O_2$) following the onset of exercise in each step condition

	AVOVA	U→M	U→VH	M→VH
$\dot{V}O_{2bl}$ (l/min)	< .001	0.69 ± 0.18	0.72 ± 0.17	$1.21 \pm 0.34^\dagger$
TD _p (s)	.01	11 ± 3	9 ± 3	$8 \pm 7^*$
$\tau\dot{V}O_{2p}$ (s)	< .001	23 ± 6	26 ± 8	$42 \pm 15^\dagger$
CI ₉₅ (s)	.006	7 ± 2	6 ± 3	$10 \pm 4^\dagger$
A _p (l/min)	< .001	0.53 ± 0.19	$1.25 \pm 0.30^*$	$0.82 \pm 0.31^\dagger$
G _p (ml/min/W)	.04	9.9 ± 1.3	$9.1 \pm 1.0^*$	9.6 ± 1.2
TD _{Sc} (s)	-	-	160 ± 33	184 ± 35
A _{Sc} (l/min)	-	-	0.21 ± 0.13	$0.11 \pm 0.06^\dagger$
$\dot{V}O_{2tot}$ (l/min)	< .001	1.22 ± 0.36	$2.18 \pm 0.55^*$	$2.14 \pm 0.58^*$
G _{tot} (ml/min/W)	.008	9.9 ± 1.3	$10.5 \pm 0.9^*$	$11.0 \pm 1.0^*$

Values are mean \pm SD. $\dot{V}O_{2bl}$, mean $\dot{V}O_2$ during baseline cycling; TD_p, phase II time delay; $\tau\dot{V}O_{2p}$, phase II time constant; CI₉₅, 95% confidence interval for $\tau\dot{V}O_{2p}$; A_p, amplitude of phase I + II, excluding $\dot{V}O_{2bl}$; TD_{Sc}, slow component time delay; A_{Sc}, amplitude of slow component; $\dot{V}O_{2tot}$, mean $\dot{V}O_2$ during the last 30 s of cycling; G_p and G_{tot}, 'gain' (i.e. $\Delta\dot{V}O_2/\Delta W$) of the phase II component and at end-exercise, respectively. Significant differences ($P < 0.05$) vs. *U→M and vs. †other step conditions.

Table 2. Kinetics of NIRS-derived variables following the onset of exercise in each step condition

	ANOVA	U→M	U→VH	M→VH
TOI _{bl} (%)	< .001	69 ± 1	69 ± 3	66 ± 4 [†]
TOI _{end} (%)	< .001	66 ± 3	58 ± 4*	58 ± 4*
TD-TOI (s)	< .001	13 ± 3	7 ± 2*	5 ± 5*
τTOI (s)	< .001	9 ± 3	7 ± 2	15 ± 5 [†]
SEE		1 ± 1	1 ± 0	1 ± 1
MRT-TOI (s)	< .001	22 ± 4	14 ± 2 [†]	20 ± 6
TD-Δdeoxy[Hb+Mb] (s)	< .001	13 ± 3	8 ± 2*	6 ± 3 [†]
τΔdeoxy[Hb+Mb] (s)	< .001	11 ± 7	9 ± 3	18 ± 6 [†]
SEE		1 ± 1	1 ± 0	1 ± 0
MRT-Δdeoxy[Hb+Mb] (s)	< .001	24 ± 5	17 ± 3 [†]	24 ± 8

Values are mean ± SD. TOI, tissue oxygenation index; Δdeoxy[Hb+Mb], change in deoxygenated haemoglobin + myoglobin concentration; MRT, mean response time; SEE, standard error of the estimate for the τTOI and τΔdeoxy[Hb+Mb]. Significant differences ($P < 0.05$) vs. *U→M and vs. [†]other step conditions.

FIGURE LEGENDS

Figure 1. Pulmonary oxygen uptake ($\dot{V}O_2$) response in a representative participant following the onset of step cycling in each condition. The vertical dashed lines indicate the onset of step exercise. The solid black lines denote the least squares regression fit of the phase II $\dot{V}O_2$ kinetic response [see Equation (1)].

Figure 2. Muscle deoxy[Hb+Mb] response of the vastus lateralis in a representative participant following the onset of step cycling in each condition. Data are normalized relative to the end-exercise amplitude after correcting for the mean value during unloaded (15 W) pedaling. The vertical dashed lines indicate the onset of step exercise. The solid black lines denote the least squares regression fit of the primary deoxy[Hb+Mb] kinetic response [see Equation (1)].

Figure 3. Tissue oxygenation index (TOI) of the vastus lateralis in a representative participant following the onset of step cycling in each condition. The vertical dashed lines indicate the onset of step exercise. The solid black lines denote the least squares regression fit of the primary TOI kinetic response [see Equation (1)].

Figure 4. Comparison of $\dot{V}O_2$ and muscle deoxygenation kinetics following the onset of step cycling. Panels A and B show the group mean \pm SD $\tau\dot{V}O_{2p}$ (*black bars*) and mean response time (MRT) of Δ deoxy[Hb+Mb] and TOI (*white bars*) within each step condition. Panels C and D present those values for $\tau\dot{V}O_{2p}$ minus the MRT- Δ deoxy[Hb+Mb] and MRT-TOI during U \rightarrow M, U \rightarrow VH and M \rightarrow VH, respectively. $^{\#}P < 0.01$ relative to the $\tau\dot{V}O_{2p}$ within condition, $^{*}P < 0.01$ vs. U \rightarrow M, and $^{\dagger}P < 0.05$ vs. other step conditions.

Figure 5. Group mean normalized ratio between the adjustment of deoxy[Hb+Mb] relative to $\dot{V}O_2$ following the onset of U \rightarrow VH (*black circles*) and M \rightarrow VH (*white circles*) step transitions. The ratio was calculated after normalizing both signals relative to the total increase (Δ) between baseline and end-exercise (i.e. 0 – 100%) with the $\dot{V}O_2$ data left shifted by 15 s to account for the muscle-to-lung transit delay. Please note error bars are excluded for clarity. Note the greater ‘overshoot’ area above unity (horizontal dashed line) within the initial few minutes of M \rightarrow VH compared to U \rightarrow VH exercise.

Figure 6. Relationship between alterations (Δ) in the $\tau\dot{V}O_{2p}$ and G_p with chronological age following the onset of work-to-work cycling transitions. The y-axis values represent those in M \rightarrow VH minus U \rightarrow M (A – B) and U \rightarrow VH (C – D), respectively, $^{*}P < 0.01$.

Figure 7. Pulmonary $\dot{V}O_2$ response during U \rightarrow M (black circles) and M \rightarrow VH (white circles) step cycling in a male youth participant aged 12 y (A – B) and 16 y (C – D) with an estimated maturity offset from PHV of -2.4 and $+2.3$ y, respectively. The $\dot{V}O_2$ data is expressed per unit change in work rate (i.e. ‘gain’). Continuous lines represent the fitted responses extrapolated backward to the pre-transition value (i.e. during the phase I region) with the model extended to 6 min during U \rightarrow M (A and C). See text for further explanation.

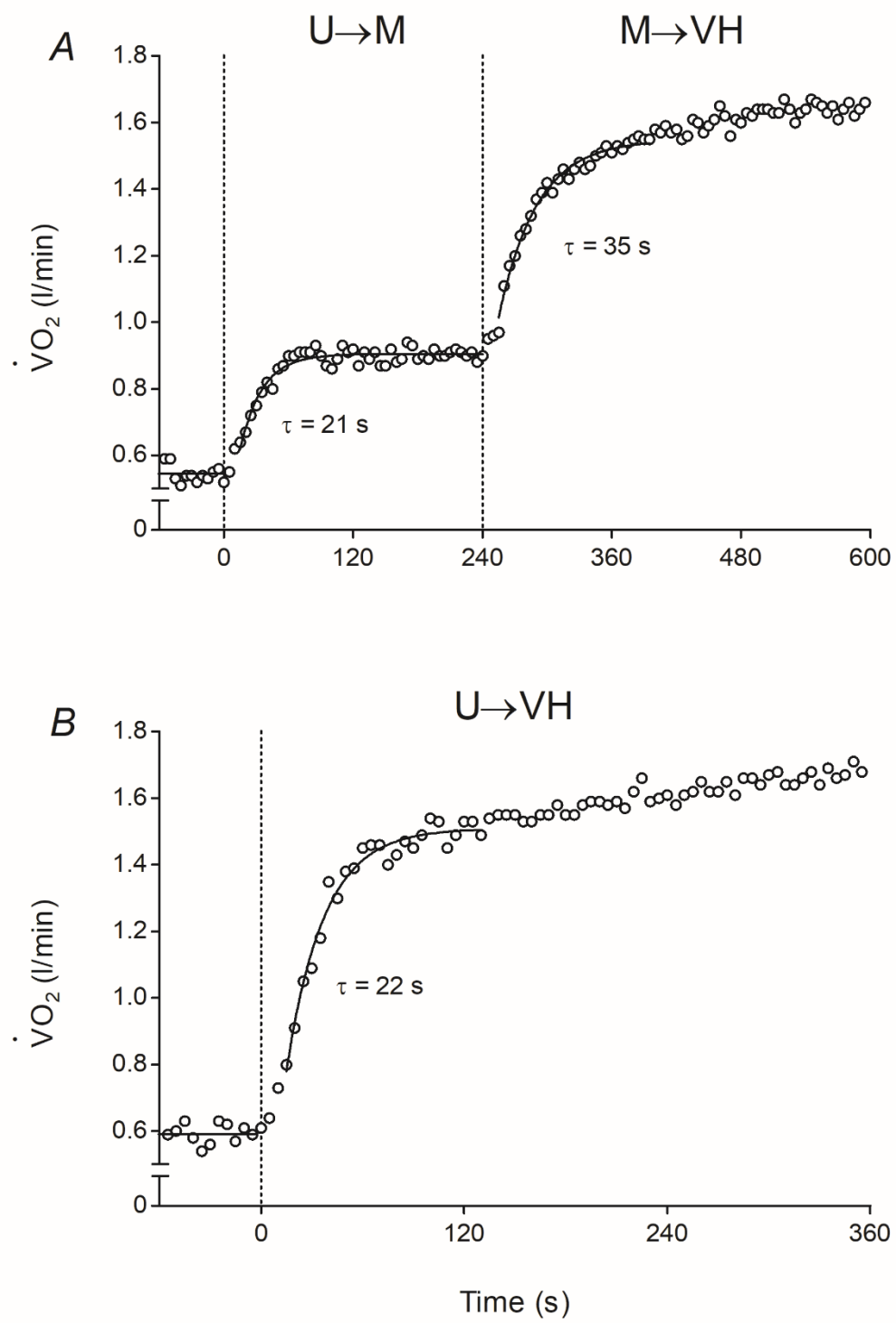


Fig. 1

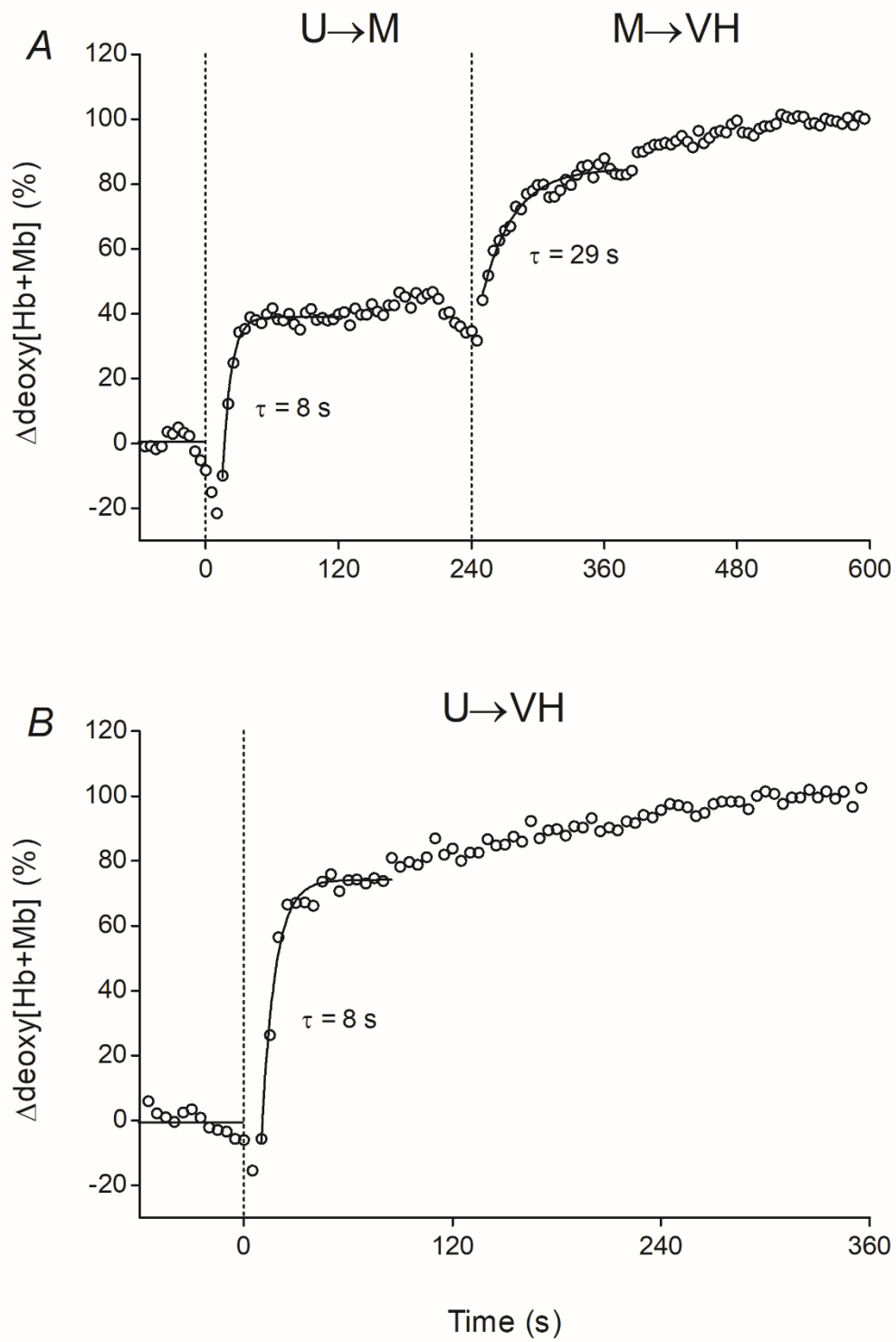


Fig. 2

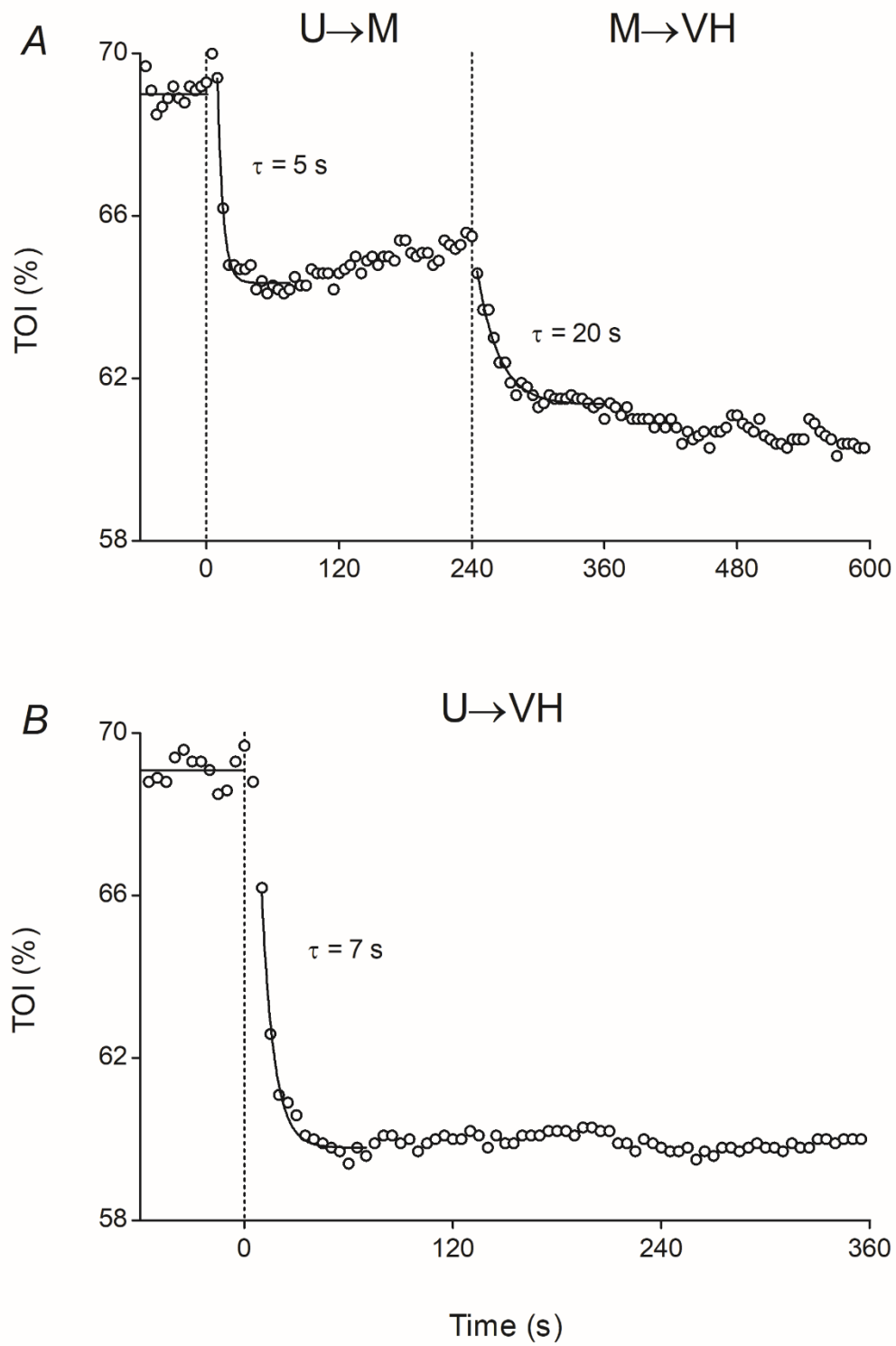


Fig. 3

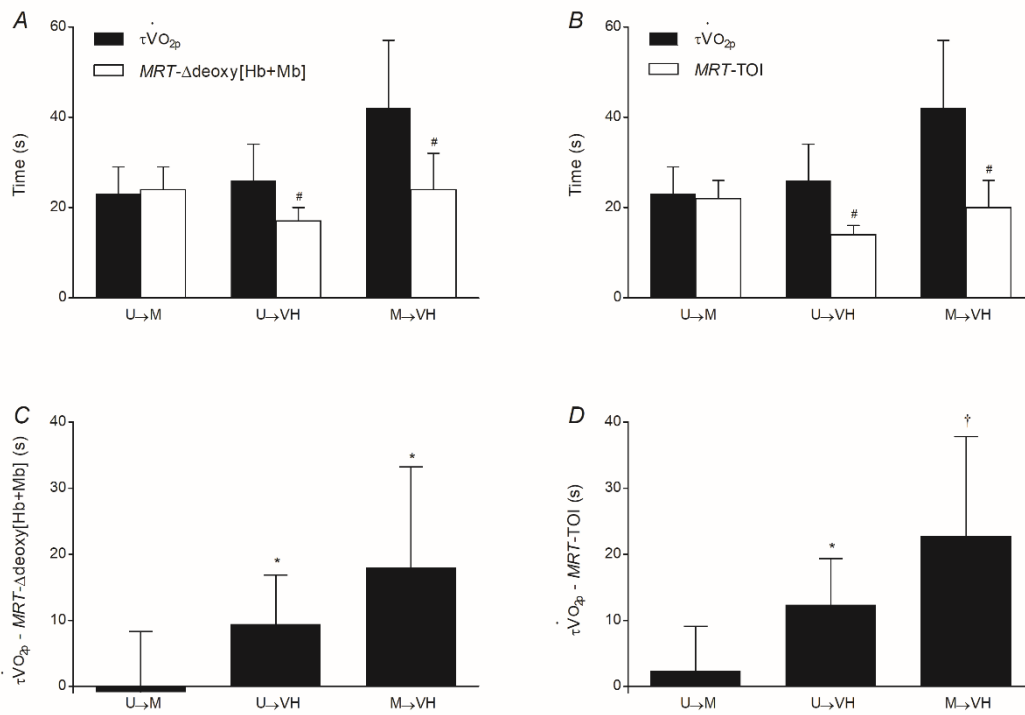


Fig. 4

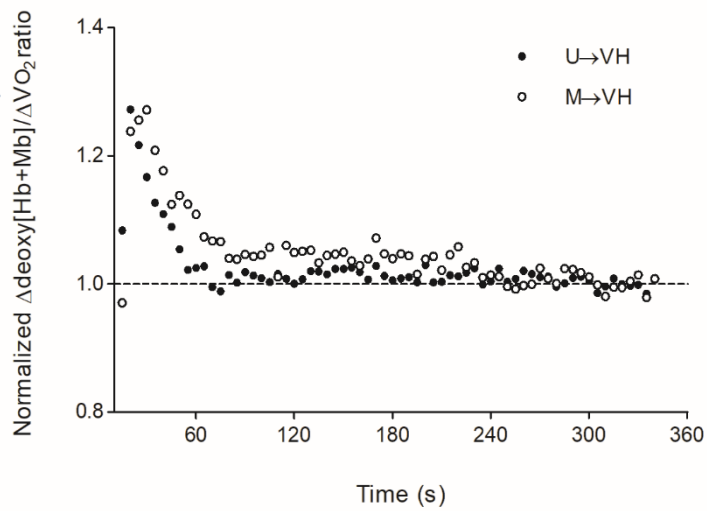
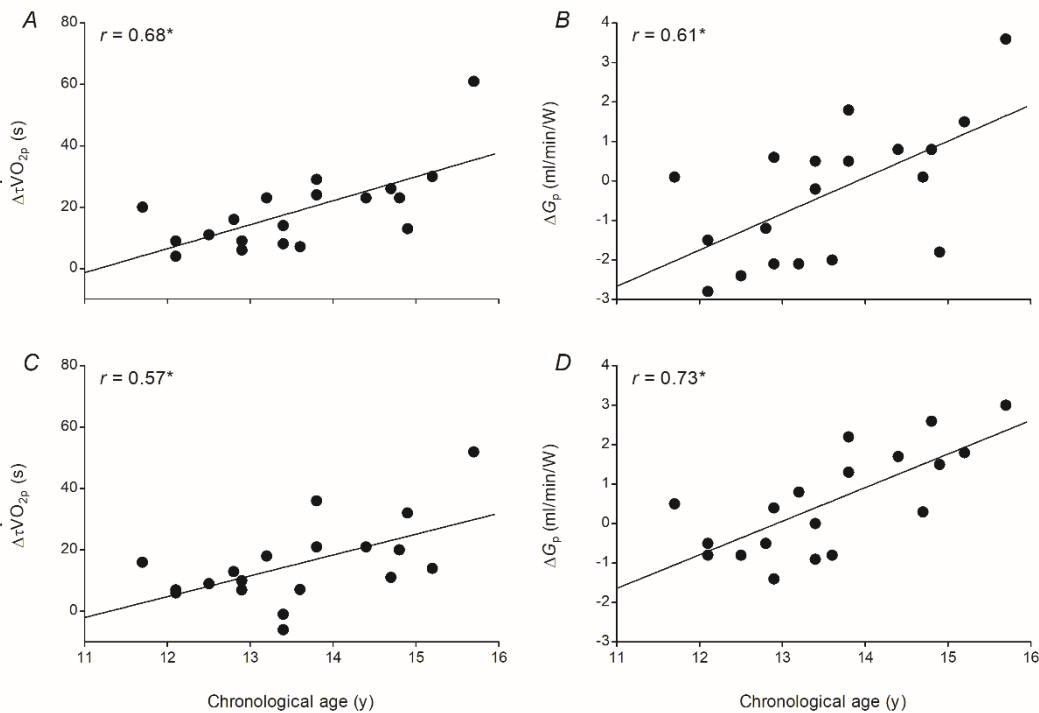


Fig. 5

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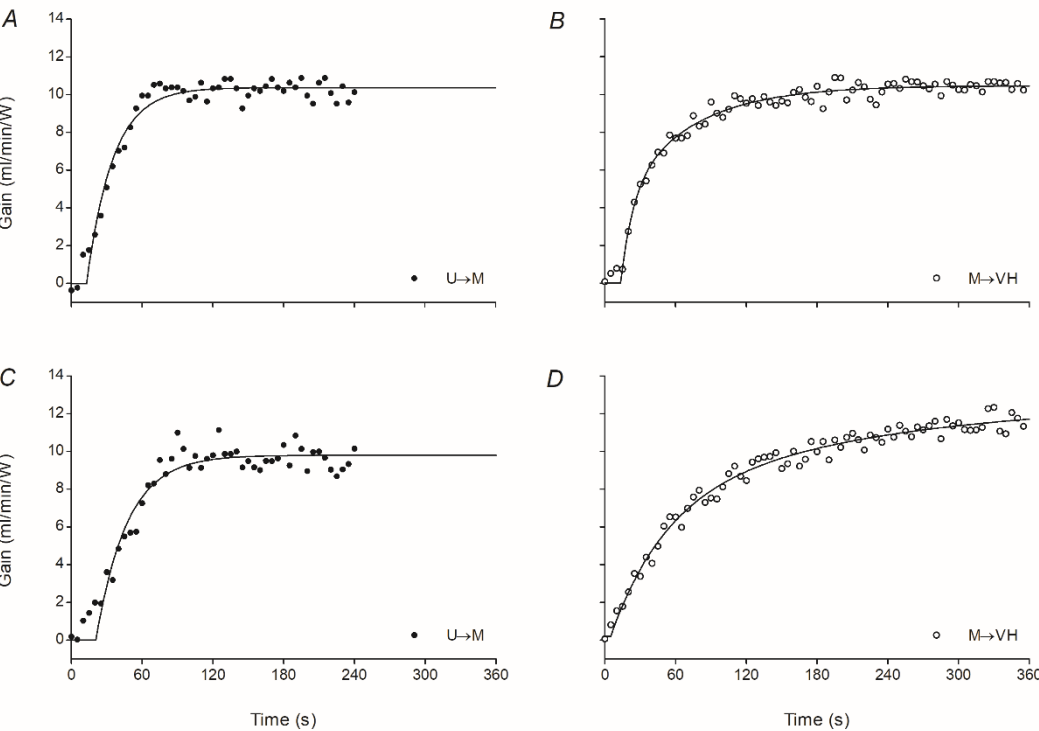
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Fig. 6



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Fig. 7